

【Product Name】 MagPure DNA Micro Kit

【Product specifications】 200 Preps/Kit

【Intended Use】

This product is suitable for rapid extraction of DNA from tissue, cells, blood, saliva, swabs, blood spots, semen and other clinical samples. DNA can be used directly for PCR, quantitative PCR, Southern Blot, test of virus DNA and so on.

【Principle】

This product is based on the purification method of high binding magnetic particles. The sample is lysed and digested under the action of lysate and Protease. DNA is released into the lysate. After adding magnetic particles and binding solution, DNA will be adsorbed on the surface of magnetic particles, and impurities such as proteins will be removed without adsorption. The adsorbed particles were washed with washing solution to remove proteins and impurities, washed with ethanol to remove salts, and finally DNA was eluted by Elution Buffer.

【Main Composition】

Cat.No.	IVD3101	Composition
MagBind Particles	4.5 ml	Magnetic Particles
Proteinase K	90 mg	Protease
Protease Dissolve Buffer	10 ml	Glycorel/Tris/CaCl ₂
Buffer ATL	60 ml	Tris/EDTA/SDS
Buffer AL	60 ml	Guanidine Salt
Buffer BD*	20 ml	sodium perchlorate
Buffer BXW1 *	110 ml	Guanidine Salt
Elution Buffer	30 ml	Tris/EDTA

【Storage conditions and Validity】

Proteinase K, MagBind Particles should be stored at 2–8°C upon arrival. However, short-term storage (up to 18 weeks) at room temperature (15–25°C) does not affect their performance. The remaining kit components can be stored dry at room temperature (15 – 25°C) and are stable for at least 18 months under these conditions.

【Preparation before Use】

- Add 4.5ml ml Protease Dissolve Buffer to the Proteinase K, and store at -20~8°C after dissolve
- Dilute Buffer BXW1 with 140ml 100% ethanol and store at room temperature

- Dilute Buffer BD with 80ml 100% ethanol and store at room temperature
- Prepare 75% Ethanol using Absolute Ethanol and store at room temperature.

【 Protocol 】

A. solid tissue (1~10mg)

1. Cut ~10mg tissue into small pieces and transfer into a new 1.5ml centrifuge tube. Add 200ul Buffer ATL and 20ul Proteinase K, Shaking at 55°C for 30~180 minutes.
2. Add 200ul Buffer AL to the samples, vortex to mix and incubation at 70°C for 10 minutes.

B. Anticoagulated blood or Plasma (200ul)

1. Transfer 20ul Proteinase K to a new 1.5ml centrifuge tube.
2. Add 200ul whole blood, plasma or other body fluids to the tube, shake to mix for 5 seconds.
3. Add 200ul Buffer AL to the samples. Inverting for 3~5 times, and then vortex at maximum speed for 10 seconds. Incubate at 70°C for 10 minutes.

C. Saliva Sample (Preserved at Orange Tube)

1. Transfer 20ul Proteinase K to 1.5ml centrifuge tube.
2. Add 300ul Saliva to the tube and shake to mix for 5 seconds. Incubate at 55°C for 30 minutes.
3. Add 100ul Buffer AL to the samples, vortex to mix.

D. Culture cells

1. Collect Cells (< 1 x 10⁶) by centrifugation at 2,000 x g for 5 min. Remove the Liquids.
2. Add 200ul Buffer PBS and 20ul Proteinase K to the sample, resuspend the cells by vortexing.
3. Add 200ul Buffer AL and vortex for 10 seconds. Incubation at 70°C for 10 minutes.

E. Semen sample

1. Transfer 100ul semen to 1.5ml centrifuge tube.
2. Add 100ul Buffer ATL, 10ul DTT Solution (1M) and 20ul Proteinase K to the samples. Shaking at 55°C for 30 minutes.
3. Add 200ul Buffer AL to the sample, then vortex to mix and incubate at 70°C for 10 minutes.

F. Swab DNA extraction

1. Transfer the swabs to the 2ml centrifuge tube.
2. Add ~500ul ATL and 20ul Proteinase K. Shaking at 55°C for 15~30 minutes.
3. Transfer 400ul of the supernatant into a new tube.

G. Blood stains/Seminal Spots

1. Transfer the 3 slices(3mm) to the 2.0ml centrifuge tube. Add 250ul Buffer ATL and 20ul Proteinase K to the sample. Shaking at high speed for 30~60min at 55°C.
2. Add 250ul Buffer AL to the samples, Shaking at high speed for 10 min at 70°C.

3. Centrifuge at 13,000 x g for 1 min. Transfer 400ul of the supernatant to a new centrifuge tube.

H. FFPE Samples

1. Using a scalpel, trim excess paraffin off the sample block. Cut up to 1~3 sections 5–10 µm thick into a 1.5 ml microcentrifuge tube. Remove Paraffin by xylene or Buffer DPS (no provided).
2. Add 200ul Buffer ATL and 20µl Proteinase K to the sample, mix well and incubate at 56°C for 60min, 90°C for 60 min.
3. Cool to room temperature, add 200ul Buffer AL and mix well.

Manul Purify by Magentic Particles.

1. **Add 20 µl MagBind Particles and 400µl Buffer BD** to the samples or the supernatant. Mix thoroughly by inverting for 15~30 times. Place the tube to the magnetic stand for 5 minutes until the beads have formed a tight pellet. Then remove the supernatant.
2. **Add 500µl Buffer BXW1** and vortex for 15 seconds to re-suspend beads. Place the tube to the magnetic stand for 2 minute until the beads have form a tight pellet. Then remove the supernatant.
3. **Add 500µl 75% ethanol**, and vortex for 15 seconds to re-suspend beads. Place the tube to the magnetic stand for 2 minute until the beads have form a tight pellet. Then remove the supernatant.
4. **Add 500µl 75% ethanol**, and vortex for 15 seconds to re-suspend beads. Place the tube to the magnetic stand for 2 minute until the beads have form a tight pellet. Then remove the supernatant.
5. Centrifuge shortly to collect liquid on the tube. Place the tube to the magnetic stand and remove all the liquid carefully. Air dry for 6 minutes.
6. **Add 30~100µl Elution Buffer to the sample**, re-suspend the beads by vortex. Incubate at 55°C for 10 minutes by shaking. If there is no shaking device, vortex 2~3 times to mix .
7. Place the tube to the magnetic rack for 2 minutes.
8. Transfer the supernatant containing the purified DNA to a clean 1.5ml centrifuge tube.

Auto Purify by KingFisher Flex

1. Add the Reagents/sample to the wells of the deep well plate according to the table below.













Name of the Plate	Pre-loaded reagents	Addition before use
Sample plate	400µl Buffer BD, 20µl MagBind Particle	400µl Lysis Sample or Supernatant
Wash Plate 1	500µl Buffer BXW1, Put in 96 magnetic Tip	
Wash Plate 2	500µl 75% ethanol	
Wash Plate 3	500µl 75% ethanol	
Elution plate	100µl Elution Buffer	

2. Turn on the machine, start the corresponding program(IVD3101_F_96CE).
3. Place the 96-well plate into the instrument as prompted.
4. Finish the operation after ~30 minutes.
5. Remove the 96-well plate and magnetic jacket.
6. Store the Elute product at -20~8°C.

【Basic Information】

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【Explanation of Marks】

	The product is used in vitro, please don't swallow		Please don't reuse it
	Validity		Please read the instruction book carefully before using
	Warning, please refer to the instructions in the annex		Manufacturer
	Temperature scope within which the product is reserve		Batch number
	European union authorization representativ		Keep dry
	Avoid overexposure to the sun		Don't use the product when the package is damaged